

Review

## Advances in the Molecular Biology of Malignant Mesothelioma

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**Malignant mesothelioma (MM) is a highly aggressive tumor with a dismal prognosis. The incidence of MM is increasing as a result of widespread exposure to asbestos. As for the molecular alterations that occur in MM, chromosome alterations including homo-deletion of the *P16* and *P14* genes located in the 9p21 are well known. Mutations are rare in the *P53* and *Ras* genes, which are frequently present in epithelial solid tumors. However, mutations are frequently present in the neurofibromatosis type 2 gene. Epigenetic alterations including DNA methylation have been found in the MM, the profile of which is different from that of lung cancer, although differential diagnosis is sometimes clinically difficult. As in other malignant tumors, genes that are related to immortalization, proliferation, metastasis, angiogenesis, and anti-apoptosis are also overexpressed in MM, contributing to its malignant phenotype. It is of interest that simian virus 40 has been implicated to be one of the causative factors of MM in western countries. Although the causative role of asbestos is well-known in MM, much less information is available for MM than for other malignant tumors regarding the molecular alterations that occur in the disease. In terms of future tasks, it will be necessary to apply the knowledge that is learned about molecular alterations to clinical practice and to further elucidate the pathogenesis of MM with extensive research.**

**Key words:** malignant mesothelioma, P16, methylation

**M**alignant mesothelioma (MM) is an aggressive tumor that develops from the pleura or other mesothelial surfaces and is strongly associated with exposure to asbestos. Many MMs have already progressed by the time they are discovered and show an extremely poor prognosis [1]. Multimodality therapy centered on surgical resection is indicated for the

treatment of MM in the early stage, but so many cases are found at advanced stage. No method of treatment, including chemotherapy or radiotherapy, has yet been established for advanced cases. Thus, new strategies for the diagnosis and treatment of MM are needed. In fact, much less information regarding molecular biological alterations is available for MM than for other solid neoplasms, even though an understanding of their molecular biological characteristics is needed to establish diagnostic, treatment, and prevention methods. This review summarizes the representative molecular biological alterations in MM that have been

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demonstrated thus far.

### Molecular Alterations in MM

**Effect of asbestos.** Asbestos, especially amphiboles fibers with a high length-to-diameter ratio, is a well-established carcinogen of MM [2, 3]. Research on the molecular alterations produced by asbestos in mesothelial cells has been under way since the 1990s. There may be a 35- to 40-year latent period before the development of asbestos-induced MM. The clinical manifestations of MM are thought to arise as a result of a build-up of many molecular alterations. Asbestos fibers show increased expression of the proto-oncogenes *c-fos* and *c-jun* [4]. These proteins are localized in the nucleus, resulting in cell proliferation and transcription, which are thought to be the initial intranuclear alterations caused by asbestos. Asbestos in nature also has a cytotoxic effect on mesothelial cells *in vitro*. On the other hand, asbestos also promotes secretion of the pro-inflammatory cytokine TNF- $\alpha$  in mesothelial cells and macrophages, leading to the activation of NF- $\kappa$ B through promoting the expression of the TNF- $\alpha$  receptor in mesothelial cells [5, 6]. Because NF- $\kappa$ B plays a role in cell proliferation and anti-apoptosis, MM cells are assumed to undergo neoplastic transformation as a result of the activation of the NF- $\kappa$ B pathway [5].

**Chromosome alterations.** Numerous alterations at the chromosome level have been reported in MM based on the results of comparative genomic hybridization and loss of homozygosity (LOH) analyses. While both chromosome amplifications and deletions have been found, deletions are more frequent than amplifications, with more involved sites. There have been many publications regarding the alterations of chromosome loci. The most frequent deletions and the chromosome locus amplifications that have been published are shown in Table 1 [7–17]. A particularly high frequency of homo-deletions is seen in the 9p21 region in MM, thus causing a high frequency of deletions of the *P16* and *P14* genes that are located on 9p21 and a loss of expression of their proteins [18]. Indeed, an attempt is being made to apply this finding for diagnosis by using the fluorescence *in situ* hybridization method to identify homo-deletion of the *P16* gene in MM cells in pleural fluid [19]. As a potential therapeutic strategy, cell cycle arrest is induced in

MM cells to which either p16 or p14 expression constructs have been introduced [20, 21]. The mechanism of the *P16* alterations and malignant transformation has been found to involve the loss of p16 protein, which causes a breakdown of the cell-cycle control mechanism by inhibiting the phosphorylation of retinoblastoma (Rb) protein, which controls the cell cycle. In addition, the loss of p14 protein results in the activation of mdm<sup>2</sup> protein, a p53 ubiquitin ligase, and this activation is thought to be linked to the destabilization of p53 protein, thus causing alterations in cell cycle control [22] (Fig. 1). Deletion of *P16* is significantly more common in MMs that have a sarcomatous component and is considered to be a negative prognostic factor for MM independent of the sarcomatoid type [23].

Differences in chromosome alterations have also been reported in the histological subtypes, which may contribute to the biological and morphological differences among subtypes [13, 24].

**Gene mutations.** Neurofibromatosis type 2 (*NF2*) gene mutations have been discovered in MM [25]. The *NF2* gene is originally known to be the causative gene in neurofibroma (type II). *NF2* gene mutations have been observed in approximately 50% of MM cases. There has been a report of LOH of 22q12, where the *NF2* gene is located, in almost 100% of mutant cases [26]. Because *NF2* gene mutations are not observed in lung cancer, the presence of these mutations is considered useful for making the

**Table 1** Representative chromosomal abnormalities in MM

Chromosomal abnormalities		Related genes
Loss	Gain	
1p21–22 3p21 4p, 4q		<i>RASSF1A</i>
	5p	
	6q	
	8q	<i>P16, P14</i> <i>RB</i>
9p21 13q13–14		
14q 15q 17p12		<i>P53</i>
	17q	
22q12		<i>NF2</i>

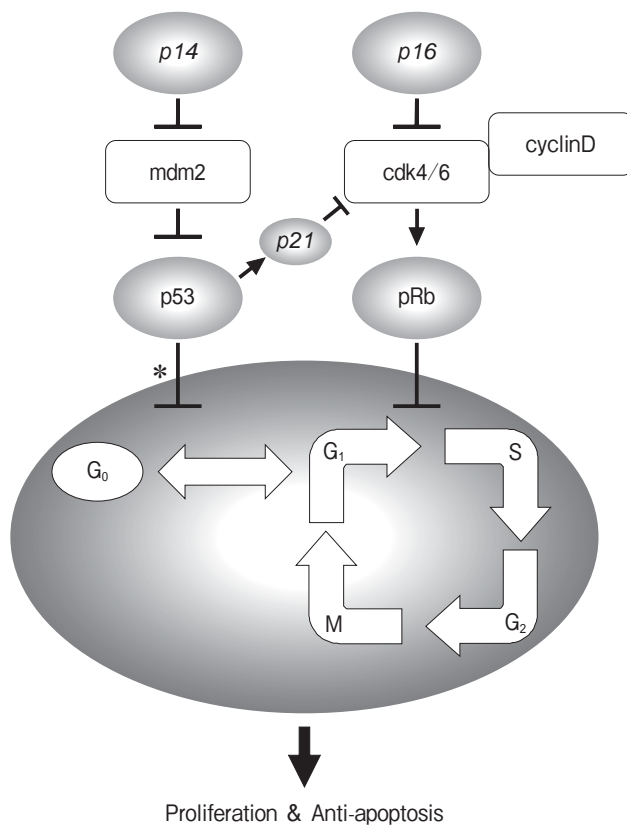
differential diagnosis from lung cancer, especially from adenocarcinoma of the lung [25, 27, 28]. However, since there are no hot spots at the mutation sites in the *NF2* gene, it is necessary to examine all 17 exons in order to detect mutations, as it is not realistic to use *NF2* gene mutations as a marker for diagnosis at the present time. When a simple method to detect *NF2* gene mutations is developed, the *NF2* gene is likely to become a valuable diagnostic marker. While mutations in the Wilms' tumor 1 (*WT1*) gene have also been found, their frequency is low, and thus it is difficult to use them for molecular diagnosis [29, 30]. Mutations in other genes including the *P53*, *Ras*, and *RB* genes, which have been found at high frequencies in other malignant tumors, are very rare [31–33]. It is assumed that alterations in the *P53* and *RB* genes in particular are unnecessary, because even if

the *P53* and *RB* genes are of the wild type, they do not function properly because of the *P14* and *P16* genes deletions as described above. However, the reason that the DNA sequences of *P53* and *RB* are intact in MM is unclear. This may be a crucial key to understanding the pathogenesis of MM.

**DNA methylation.** Gene inactivation by epigenetic alteration has been established as a crucial mechanism that satisfies Knudson's hypothesis that both alleles of a tumor suppressor gene (TSG) must be inactivated for carcinogenesis. Promoter methylation and the associated event of histone deacetylation are epigenetic changes in chromatin structure that cause gene silencing without altering the DNA sequence [34, 35]. Methylation of TSGs, including methylation of the *RASSF1A* gene, has been observed in MM as well, strongly suggesting that methylation of the promoter region of TSGs contributes to the neoplastic transformation and progression of MM [36–39]. Since the methylation profile varies with the histological subtype of the MMs, the mechanism of neoplastic transformation may differ according to the histological subtype [37]. The methylation profile of MM is clearly different from those of adenocarcinoma of the lung, and thus it can be used as a diagnostic tool [37]. Because the histological findings of these 2 diseases are sometimes too similar to distinguish them, the methylation profile along with *NF2* mutation status can be useful for performing differential diagnosis.

Most of the studies of methylation in MM have thus far been conducted on specimens in the United States, where SV40 infection is suspected to be one of the causes. Because the methylation profile of some genes differs between SV40-infected and non-infected MMs, the methylation profile of MM in counties like Japan where there is no SV40 involvement may be totally different from that in the United States. Our previous study indicates that methylation of the insulin-like growth factor-binding protein-3 (*IGFBP-3*) gene, which is thought to control IGF function by suppressing the IGF-1 receptor, has been shown to be more frequent in MM in Japan than in the United States. This finding suggests the presence of racial or regional differences in the genes that undergo methylation in MM [40].

**Activation of telomerase.** Telomerase activation, which is thought to be one of the causes of immortalization, is increased in malignant tumor cells.



**Fig. 1** Schema showing the role of p16 and p14 in the cell cycle. The loss of p16 and p14 function leads to alterations in cell cycle control, resulting in cell proliferation and anti-apoptosis. \*G<sub>0</sub> arrest by p53 pathway mainly occurs when DNA is damaged.

It is also known to be high in MM [41]. The expression of hTERT, which is closely associated with telomerase activity, has been found in 90% of MM cases. Since its expression is not observed in mesothelial cells in non-tumorous portions, it may be capable of serving as a marker for the malignant transformation of mesothelial cells [41].

**Cell proliferation.** An increase in cell proliferation is seen in MM as in other malignant tumors. Cell proliferation increases particularly as a result of the autocrine and paracrine function of growth factors: epidermal growth factor (EGF) [42], hepatocyte growth factor (HGF) [43], platelet-derived growth factor (PDGF) [44], transforming growth factor  $\beta$  [45], IGF-1, and IGF-II [46]. At the same time, high levels of expression of the receptors for these growth factors such as EGFR, c-MET, and IGF-R1 have been found. Moreover, alterations of the Wnt signaling system, which is involved in the proliferation and generation of normal cells, have recently been reported in MM [47].

**Invasiveness and angiogenesis.** MMs are highly locally invasive tumors but distant metastasis also sometimes occurs in advanced cases. The matrix metalloproteinases (MMPs) MMP-2 and MMP-9, in particular, are known to be related to the invasion and metastasis of MM. MMP-2 is also considered to be a negative prognostic factor [48]. It has also been suggested that the microvessel density of tumors is a prognostic indicator [49], and that the vascular endothelial growth factor that contributes to angiogenesis is overexpressed [50].

**Anti-apoptosis.** The Bcl-2 family of proteins is known to be involved in apoptosis and strong expression of these protein is seen in many malignant tumors. Whereas the frequency of expression of Bcl-2 protein itself in MM is low, strong expression of Bcl-XL (a member of the Bcl-2 family) and Bax, which have potent anti-apoptotic potential, has been found [51–53]. Strong expression of the inhibitor of apoptosis protein (IAP) and survivin, which are considered resistance factors for chemotherapy, has also been observed [54].

**Association with SV40.** There is a hypothesis in western countries that infection with SV40, a virus whose original hosts are monkeys, is one of the causative factors of MM [55, 56]. In support of the hypothesis, SV40 large T antigen DNA has been

detected in approximately half of MM cases in the United States [57]. The reason why SV40 was detected in human tissue has been presumed to be that the polio vaccine produced between 1955 and 1963 was contaminated with SV40 in the manufacturing process, and that SV40 infected humans through inoculation with the contaminated vaccine. There have been many arguments against this explanation, and this topic continues to invite debate [58]. There is concern that the vaccine that may have been contaminated was used in Japan from 1961 to 1963, but the result of a study in Japan regarding the association between MM and SV40 was negative [59]. Inactivation of p53 and Rb protein by SV40 large T antigen has been demonstrated as the mechanism of carcinogenesis by SV40 [60]. Moreover, in human mesothelial cells that were infected with SV40 and immortalized, an increase in hTERT activity was observed in the early stage, and methylation and a decrease in the expression of the *RASSF1A* gene was observed in the late stage, both of which are assumed to be related to SV40-associated MM [61, 62]. Malignant transformation of mesothelial cells by SV40 is thought to be promoted by its combination with the presence of asbestos [60].

**Attempts at comprehensive analysis of expression.** The differential display and microarray methods are useful assays for the comprehensive analysis of mRNA expression. Attempts have been made to identify altered gene expression levels through the comprehensive analysis of mRNA expression in MM tissue and in non-tumorous tissue. Alterations in the level of variety gene expression, including the activation of apoptosis-associated genes and gene groups involved in glucose metabolism, mRNA transcription, and cytoskeleton formation, have been occasionally reported [63, 64] but not universally identified. Although some studies have also explored the possibility of identifying prognostic factors by expression profiling, they have not reached the clinical application stage and require further investigation [65, 66].

## Concluding Remarks

The worldwide incidence of MM is considered to be increasing [1], especially in Japan and non-western countries, where asbestos continued to be heavily used

after its use had been discontinued in western countries. Thus, the urgent establishment of preventive, diagnostic and treatment strategies for MM is mandatory. Also, the roles of erionite and of genetic factors, as shown by the outbreak in certain areas of Caddadochia, Turkey, can be crucial clues for understanding the pathogenesis of MM [67]. So far, extensive research has made some of the critical molecular alterations clear, but the findings are still not adequately reflected in clinical settings. In terms of future tasks, it will be necessary to apply what has been learned so far to clinical practice and to further elucidate the diseases using a molecular biological approach.

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## References

- Robinson BW and Lake RA: Advances in malignant mesothelioma. *N Engl J Med* (2005) 353: 1591–1603.
- Wagner JC, Sleggs CA and Marchand P: Diffuse pleural mesothelioma and asbestos exposure in the North Western Cape Province. *Br J Ind Med* (1960) 17: 260–271.
- Pass HI, Mitchell JB, Johnson DH, Turrisi AT and Minna JD: Lung cancer: principle and practice. 2nd Ed, Lippincott Williams and Wilkins, Philadelphia (2000) pp 375–378.
- Heintz NH, Janssen YM and Mossman BT: Persistent induction of c-fos and c-jun expression by asbestos. *Proc Natl Acad Sci USA* (1993) 90: 3299–3303.
- Yang H, Bocchetta M, Kroczyńska B, Elmishad AG, Chen Y, Liu Z, Bubici C, Mossman BT, Pass HI, Testa JR, Franzoso G and Carbone M: TNF- $\alpha$  inhibits asbestos-induced cytotoxicity via a NF- $\kappa$ B-dependent pathway, a possible mechanism for asbestos-induced oncogenesis. *Proc Natl Acad Sci USA* (2006) 103: 10397–10402.
- Philip M, Rowley DA and Schreiber H: Inflammation as a tumor promoter in cancer induction. *Semin Cancer Biol* (2004) 14: 433–439.
- Flejtner WL, Li FP, Antman KH and Testa JR: Recurring loss involving chromosomes 1, 3, and 22 in malignant mesothelioma: possible sites of tumor suppressor genes. *Genes Chromosomes Cancer* (1989) 1: 148–154.
- Fletcher JA, Kozakewich HP, Hoffer FA, Lage JM, Weidner N, Tepper R, Pinkus GS, Morton CC and Corson JM: Diagnostic relevance of clonal cytogenetic aberrations in malignant soft-tissue tumors. *N Engl J Med* (1991) 324: 436–442.
- Hagemeyer A, Versnel MA, Van Drunen E, Moret M, Bouts MJ, van der Kwast TH and Hoogsteden HC: Cytogenetic analysis of malignant mesothelioma. *Cancer Genet Cytogenet* (1990) 47: 1–28.
- Balsara BR, Bell DW, Sonoda G, De Rienzo A, du Manoir S, Jhanwar SC and Testa JR: Comparative genomic hybridization and loss of heterozygosity analyses identify a common region of deletion at 15q11.1–15 in human malignant mesothelioma. *Cancer Res* (1999) 59: 450–454.
- Bjorkqvist AM, Tammilehto L, Anttila S, Mattson K and Knuutila S: Recurrent DNA copy number changes in 1q, 4q, 6q, 9p, 13q, 14q and 22q detected by comparative genomic hybridization in malignant mesothelioma. *Br J Cancer* (1997) 75: 523–527.
- Bjorkqvist AM, Tammilehto L, Nordling S, Nurminen M, Anttila S, Mattson K and Knuutila S: Comparison of DNA copy number changes in malignant mesothelioma, adenocarcinoma and large-cell anaplastic carcinoma of the lung. *Br J Cancer* (1998) 77: 260–269.
- Krismann M, Muller KM, Jaworska M and Johnen G: Molecular cytogenetic differences between histological subtypes of malignant mesotheliomas. DNA cytometry and comparative genomic hybridization of 90 cases. *J Pathol* (2002) 197: 363–371.
- Taguchi T, Jhanwar SC, Siegfried JM, Keller SM and Testa JR: Recurrent deletions of specific chromosomal sites in 1p, 3p, 6q, and 9p in human malignant mesothelioma. *Cancer Res* (1993) 53: 4349–4355.
- De Rienzo A, Balsara BR, Apostolou S, Jhanwar SC and Testa JR: Loss of heterozygosity analysis defines a 3-cM region of 15q commonly deleted in human malignant mesothelioma. *Oncogene* (2001) 20: 6245–6249.
- Shivapurkar N, Virmani AK, Wistuba II, Milchgrub S, Mackay B, Minna JD and Gazdar AF: Deletions of chromosome 4 at multiple sites are frequent in malignant mesothelioma and small cell lung carcinoma. *Clin Cancer Res* (1999) 5: 17–23.
- Musti M, Kettunen E, Dragonieri S, Lindholm P, Cavone D, Serio G and Knuutila S: Cytogenetic and molecular genetic changes in malignant mesothelioma. *Cancer Genet Cytogenet* (2006) 170: 9–15.
- Prins JB, Williamson KA, Kamp MM, Van Hezik EJ, Van der Kwast TH, Hagemeyer A and Versnel MA: The gene for the cyclin-dependent-kinase-4 inhibitor, CDKN2A, is preferentially deleted in malignant mesothelioma. *Int J Cancer* (1998) 75: 649–653.
- Illei PB, Ladanyi M, Rusch VW and Zakowski MF: The use of CDKN2A deletion as a diagnostic marker for malignant mesothelioma in body cavity effusions. *Cancer* (2003) 99: 51–56.
- Frizelle SP, Grim J, Zhou J, Gupta P, Curiel DT, Geradts J and Kratzke RA: Re-expression of p16INK4a in mesothelioma cells results in cell cycle arrest, cell death, tumor suppression and tumor regression. *Oncogene* (1998) 16: 3087–3095.
- Yang CT, You L, Yeh CC, Chang JW, Zhang F, McCormick F and Jablons DM: Adenovirus-mediated p14(ARF) gene transfer in human mesothelioma cells. *J Natl Cancer Inst* (2000) 92: 636–641.
- Lowe SW and Sherr CJ: Tumor suppression by Ink4a-Arf: progress and puzzles. *Curr Opin Genet Dev* (2003) 13: 77–83.
- Lopez-Rios F, Chuai S, Flores R, Shimizu S, Ohno T, Wakahara K, Illei PB, Hussain S, Krug L, Zakowski MF, Rusch V, Olshen AB and Ladanyi M: Global gene expression profiling of pleural mesotheliomas: overexpression of aurora kinases and P16/CDKN2A deletion as prognostic factors and critical evaluation of microarray-based prognostic prediction. *Cancer Res* (2006) 66: 2970–2979.
- Knuutila A, Jee KJ, Taskinen E, Wolff H, Knuutila S and Anttila S: Spindle cell tumours of the pleura: a clinical, histological and comparative genomic hybridization analysis of 14 cases. *Virchows Arch* (2006) 448: 135–141.

25. Sekido Y, Pass HI, Bader S, Mew DJ, Christman MF, Gazdar AF and Minna JD: Neurofibromatosis type 2 (NF2) gene is somatically mutated in mesothelioma but not in lung cancer. *Cancer Res* (1995) 55: 1227–1231.
26. Cheng JQ, Lee WC, Klein MA, Cheng GZ, Jhanwar SC and Testa JR: Frequent mutations of NF2 and allelic loss from chromosome band 22q12 in malignant mesothelioma: evidence for a two-hit mechanism of NF2 inactivation. *Genes Chromosomes Cancer* (1999) 24: 238–242.
27. Bianchi AB, Mitsunaga SI, Cheng JQ, Klein WM, Jhanwar SC, Seizinger B, Kley N, Klein-Szanto AJ and Testa JR: High frequency of inactivating mutations in the neurofibromatosis type 2 gene (NF2) in primary malignant mesotheliomas. *Proc Natl Acad Sci USA* (1995) 92: 10854–10858.
28. Schipper H, Papp T, Johnen G, Pemsel H, Bastrop R, Muller KM, Wiethage T, Jaworska M, Krismann M, Schiffmann D and Rahman Q: Mutational analysis of the NF2 tumour suppressor gene in three subtypes of primary human malignant mesotheliomas. *Int J Oncol* (2003) 22: 1009–1017.
29. Kumar-Singh S, Segers K, Rodeck U, Backhovens H, Bogers J, Weyler J, Van Broeckhoven C and Van Marck E: WT1 mutation in malignant mesothelioma and WT1 immunoreactivity in relation to p53 and growth factor receptor expression, cell-type transition, and prognosis. *J Pathol* (1997) 181: 67–74.
30. Park S, Schalling M, Bernard A, Maheswaran S, Shipley GC, Roberts D, Fletcher J, Shipman R, Rheinwald J, Demetri G, et al: The Wilms tumour gene WT1 is expressed in murine mesoderm-derived tissues and mutated in a human mesothelioma. *Nat Genet* (1993) 4: 415–420.
31. Kratzke RA, Otterson GA, Lincoln CE, Ewing S, Oie H, Geradts J and Kaye FJ: Immunohistochemical analysis of the p16INK4 cyclin-dependent kinase inhibitor in malignant mesothelioma. *J Natl Cancer Inst* (1995) 87: 1870–1875.
32. Papp T, Schipper H, Pemsel H, Bastrop R, Muller KM, Wiethage T, Weiss DG, Dopp E, Schiffmann D and Rahman Q: Mutational analysis of N-ras, p53, p16INK4a, p14ARF and CDK4 genes in primary human malignant mesotheliomas. *Int J Oncol* (2001) 18: 425–433.
33. Papp T, Schipper H, Pemsel H, Unverricht M, Muller KM, Wiethage T, Schiffmann D and Rahman Q: Mutational analysis of the PTEN/MMAC1 tumour suppressor gene in primary human malignant mesotheliomas. *Oncol Rep* (2001) 8: 1375–1379.
34. Bird AP and Wolffe AP: Methylation-induced repression—belts, braces, and chromatin. *Cell* (1999) 99: 451–454.
35. Merlo A, Herman JG, Mao L, Lee DJ, Gabrielson E, Burger PC, Baylin SB and Sidransky D: 5' CpG island methylation is associated with transcriptional silencing of the tumour suppressor p16/CDKN2/MTS1 in human cancers. *Nat Med* (1995) 1: 686–692.
36. Murthy SS, Shen T, De Rienzo A, Lee WC, Ferriola PC, Jhanwar SC, Mossman BT, Filmus J and Testa JR: Expression of GPC3, an X-linked recessive overgrowth gene, is silenced in malignant mesothelioma. *Oncogene* (2000) 19: 410–416.
37. Toyooka S, Pass HI, Shivapurkar N, Fukuyama Y, Maruyama R, Toyooka KO, Gilcrease M, Farinas A, Minna JD and Gazdar AF: Aberrant methylation and simian virus 40 tag sequences in malignant mesothelioma. *Cancer Res* (2001) 61: 5727–5730.
38. Suzuki M, Toyooka S, Shivapurkar N, Shigematsu H, Miyajima K, Takahashi T, Stastny V, Zern AL, Fujisawa T, Pass HI, Carbone M and Gazdar AF: Aberrant methylation profile of human malignant mesotheliomas and its relationship to SV40 infection. *Oncogene* (2005) 24: 1302–1308.
39. Batra S, Shi Y, Kuchenbecker KM, He B, Reguart N, Mikami I, You L, Xu Z, Lin YC, Clement G and Jablons DM: Wnt inhibitory factor-1, a Wnt antagonist, is silenced by promoter hypermethylation in malignant pleural mesothelioma. *Biochem Biophys Res Commun* (2006) 342: 1228–1232.
40. Tomii K, Tsukuda K, Toyooka S, Dote H, Hanafusa T, Asano H, Naitou M, Doihara H, Kishimoto T, Katayama H, Pass HI, Date H and Shimizu N: Aberrant promoter methylation of insulin-like growth factor binding protein-3 gene in human cancers. *Int J Cancer* (2007) 120: 566–573.
41. Dhaene K, Hubner R, Kumar-Singh S, Weyn B and Van Marck E: Telomerase activity in human pleural mesothelioma. *Thorax* (1998) 53: 915–918.
42. Dazzi H, Hasleton PS, Thatcher N, Wilkes S, Swindell R and Chatterjee AK: Malignant pleural mesothelioma and epidermal growth factor receptor (EGF-R). Relationship of EGF-R with histology and survival using fixed paraffin embedded tissue and the F4, monoclonal antibody. *Br J Cancer* (1990) 61: 924–926.
43. Tolnay E, Kuhnen C, Wiethage T, Konig JE, Voss B and Muller KM: Hepatocyte growth factor/scatter factor and its receptor c-Met are overexpressed and associated with an increased microvessel density in malignant pleural mesothelioma. *J Cancer Res Clin Oncol* (1998) 124: 291–296.
44. Versnel MA, Claesson-Welsh L, Hammacher A, Bouts MJ, van der Kwast TH, Eriksson A, Willemsen R, Weima SM, Hoogsteden HC, Hagemeyer A, et al: Human malignant mesothelioma cell lines express PDGF beta-receptors whereas cultured normal mesothelial cells express predominantly PDGF alpha-receptors. *Oncogene* (1991) 6: 2005–2011.
45. Fitzpatrick DR, Bielefeldt-Ohmann H, Himbeck RP, Jarnicki AG, Marzo AL and Robinson BW: Transforming growth factor-beta: antisense RNA-mediated inhibition affects anchorage-independent growth, tumorigenicity and tumor-infiltrating T-cells in malignant mesothelioma. *Growth Factors* (1994) 11: 29–44.
46. Hodzic D, Delacroix L, Willemsen P, Bensbaho K, Collette J, Broux R, Lefebvre P, Legros JJ, Grooteclaes M and Winkler R: Characterization of the IGF system and analysis of the possible molecular mechanisms leading to IGF-II overexpression in a mesothelioma. *Horm Metab Res* (1997) 29: 549–555.
47. Lee AY, He B, You L, Dadfarmay S, Xu Z, Mazieres J, Mikami I, McCormick F and Jablons DM: Expression of the secreted frizzled-related protein gene family is downregulated in human mesothelioma. *Oncogene* (2004) 23: 6672–6676.
48. Edwards JG, McLaren J, Jones JL, Waller DA and O'Byrne KJ: Matrix metalloproteinases 2 and 9 (gelatinases A and B) expression in malignant mesothelioma and benign pleura. *Br J Cancer* (2003) 88: 1553–1559.
49. Edwards JG, Swinson DE, Jones JL, Muller S, Waller DA and O'Byrne KJ: Tumor necrosis correlates with angiogenesis and is a predictor of poor prognosis in malignant mesothelioma. *Chest* (2003) 124: 1916–1923.
50. Masood R, Kundra A, Zhu S, Xia G, Scalia P, Smith DL and Gill PS: Malignant mesothelioma growth inhibition by agents that target the VEGF and VEGF-C autocrine loops. *Int J Cancer* (2003) 104: 603–610.
51. Segers K, Ramael M, Singh SK, Weyler J, Van Meerbeeck J, Vermeire P and Van Marck E: Immunoreactivity for bcl-2 protein in malignant mesothelioma and non-neoplastic mesothelium. *Virchows Arch* (1994) 424: 631–634.
52. Narasimhan SR, Yang L, Gerwin BI and Broaddus VC: Resistance of pleural mesothelioma cell lines to apoptosis: relation to expres-

- sion of Bcl-2 and Bax. *Am J Physiol* (1998) 275: L165–171.
53. Soini Y, Kinnula V, Kaarteenaho-Wiik R, Kurttila E, Linnainmaa K and Paakko P: Apoptosis and expression of apoptosis regulating proteins bcl-2, mcl-1, bcl-X, and bax in malignant mesothelioma. *Clin Cancer Res* (1999) 5: 3508–3515.
  54. Kleinberg L, Lie AK, Florenes VA, Nesland JM and Davidson B: Expression of inhibitor-of-apoptosis protein family members in malignant mesothelioma. *Hum Pathol* (2007) 38: 986–994.
  55. Carbone M, Rizzo P, Grimley PM, Procopio A, Mew DJ, Shridhar V, de Bartolomeis A, Esposito V, Giuliano MT, Steinberg SM, Levine AS, Giordano A and Pass HI: Simian virus-40 large-T antigen binds p53 in human mesotheliomas. *Nat Med* (1997) 3: 908–912.
  56. Gazdar AF, Butel JS and Carbone M: SV40 and human tumours: myth, association or causality? *Nat Rev Cancer* (2002) 2: 957–964.
  57. Shivapurkar N, Wiethage T, Wistuba II, Salomon E, Milchgrub S, Muller KM, Churg A, Pass H and Gazdar AF: Presence of simian virus 40 sequences in malignant mesotheliomas and mesothelial cell proliferations. *J Cell Biochem* (1999) 76: 181–188.
  58. Engels EA, Katki HA, Nielsen NM, Winther JF, Hjalgrim H, Gjerris F, Rosenberg PS and Frisch M: Cancer incidence in Denmark following exposure to poliovirus vaccine contaminated with simian virus 40. *J Natl Cancer Inst* (2003) 95: 532–539.
  59. Aoe K, Hiraki A, Murakami T, Toyooka S, Shivapurkar N, Gazdar AF, Sueoka N, Taguchi K, Kamei T, Takeyama H, Sugi K and Kishimoto T: Infrequent existence of simian virus 40 large T antigen DNA in malignant mesothelioma in Japan. *Cancer Sci* (2006) 97: 292–295.
  60. Bocchetta M, Di Resta I, Powers A, Fresco R, Tosolini A, Testa JR, Pass HI, Rizzo P and Carbone M: Human mesothelial cells are unusually susceptible to simian virus 40-mediated transformation and asbestos cocarcinogenicity. *Proc Natl Acad Sci USA* (2000) 97: 10214–10219.
  61. Foddìs R, De Rienzo A, Broccoli D, Bocchetta M, Stekala E, Rizzo P, Tosolini A, Grobely JV, Jhanwar SC, Pass HI, Testa JR and Carbone M: SV40 infection induces telomerase activity in human mesothelial cells. *Oncogene* (2002) 21: 1434–1442.
  62. Toyooka S, Carbone M, Toyooka KO, Bocchetta M, Shivapurkar N, Minna JD and Gazdar AF: Progressive aberrant methylation of the RASSF1A gene in simian virus 40 infected human mesothelial cells. *Oncogene* (2002) 21: 4340–4344.
  63. Singhal S, Wiewrodt R, Malden LD, Amin KM, Matzie K, Friedberg J, Kucharczuk JC, Litzky LA, Johnson SW, Kaiser LR and Albelda SM: Gene expression profiling of malignant mesothelioma. *Clin Cancer Res* (2003) 9: 3080–3097.
  64. Gordon GJ, Rockwell GN, Jensen RV, Rheinwald JG, Glickman JN, Aronson JP, Pottorf BJ, Nitz MD, Richards WG, Sugarbaker DJ and Bueno R: Identification of novel candidate oncogenes and tumor suppressors in malignant pleural mesothelioma using large-scale transcriptional profiling. *Am J Pathol* (2005) 166: 1827–1840.
  65. Pass HI, Liu Z, Wali A, Bueno R, Land S, Lott D, Siddiq F, Lonardo F, Carbone M and Draghici S: Gene expression profiles predict survival and progression of pleural mesothelioma. *Clin Cancer Res* (2004) 10: 849–859.
  66. Gordon GJ, Rockwell GN, Godfrey PA, Jensen RV, Glickman JN, Yeap BY, Richards WG, Sugarbaker DJ and Bueno R: Validation of genomics-based prognostic tests in malignant pleural mesothelioma. *Clin Cancer Res* (2005) 11: 4406–4414.
  67. Carbone M, Emri S, Dogan AU, Steele I, Tuncer M, Pass HI and Baris YI: A mesothelioma epidemic in cappadocia: Scientific developments and unexpected social outcomes. *Nat Rev Cancer* (2007) 7: 147–154.