

Review

Glial Cell Line-derived Neurotrophic Factor (GDNF) Therapy for Parkinson's Disease

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Many studies using animals clarify that glial cell line-derived neurotrophic factor (GDNF) has strong neuroprotective and neurorestorative effects on dopaminergic neurons. Several pilot studies clarified the validity of continuous intraputamin GDNF infusion to patients with Parkinson's disease (PD), although a randomized controlled trial of GDNF therapy published in 2006 resulted in negative outcomes, and controversy remains about the efficacy and safety of the treatment. For a decade, our laboratory has investigated the efficacy and the most appropriate method of GDNF administration using animals, and consequently we have obtained some solid data that correspond to the results of clinical trials. In this review, we present an outline of our studies and other key studies related to GDNF, the current state of the research, problems to be overcome, and predictions regarding the use of GDNF therapy for PD in the future.

Key words: cell transplantation, clinical trial, encapsulation, gene therapy, neurodegenerative disease

Parkinson's disease (PD) is characterized by the degeneration of dopaminergic neurons in the nigrostriatal system [1, 2]. The established treatment for PD is oral L-DOPA [3, 4] and stereotaxic surgeries such as deep brain stimulation [5]. In Europe and the United States, fetal nigral cell transplantation has also been performed in PD patients. However, after 2 randomized clinical trials revealed insufficient functional recovery in older patients and delayed dyskinesia in some patients [6, 7], the momentum for exploring a new therapy for PD using stem cells or neurotrophic/growth factor increased. Stem cell therapy might be the most useful therapeutic

option for PD, because recent molecular/cell biological development enables us to make dopaminergic neurons from embryonic stem cells [8] or mesenchymal stem cells [9]. However, there are many problems in terms of safety and efficiency as well as ethical issues to be resolved; these problems should be overcome before this therapy is clinically applied. Neurotrophic/growth factor might also be useful for PD. Since Dr. Lin and her colleagues isolated glial cell line-derived neurotrophic factor (GDNF) and demonstrated that GDNF enhances the survival and morphological differentiation of dopaminergic neurons [10], many laboratory studies have examined GDNF therapy as a treatment for various diseases of the central nervous system (CNS), including stroke [11], spinal cord injury [12], traumatic brain injury [13] and neurodegenerative diseases, such as PD [14, 15] and amyotrophic lateral sclerosis (ALS) [16].

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GDNF is considered to be one of the strongest neuroprotectants for dopaminergic neurons. Based on the successful results of animal experiments, clinical trials were performed for PD patients using GDNF. In this review, recent findings with regard to GDNF therapy are described, as is the current status of GDNF therapy for PD patients.

Therapeutic Effects of GDNF on PD Model of Animals

Mechanisms of GDNF therapy. GDNF, a member of the transforming growth factor β [10], is known to demonstrate neuroprotective and neurorestorative effects on dopaminergic neurons [17–20] through a heterodimeric receptor complex consisting of a transmembrane receptor tyrosine kinase, Ret, and the ligand binding component GDNF-family receptor $\alpha 1$ (GFR $\alpha 1$) [21, 22]. Cell survival/death decisions are determined in part through the activation of a specific intracellular signaling cascade, such as the phosphatidylinositol 3 kinase pathway and the mitogen-activated protein kinase pathway [21, 23].

Key experiments using GDNF for PD model of animals. Dr. Hebb and his colleagues demonstrated that GDNF enhanced the survival of fetal rat dopaminergic neurons for the purpose of transplantation [18]. GDNF therapy by ventricular infusion using an osmotic minipump suppressed the neuronal degeneration induced by 6-OHDA [19, 24]. *In vivo* gene delivery of GDNF was also a feasible neuroprotective strategy for PD [20]. Recently the importance of transplanted cells as neurotrophic/growth factor suppliers has been re-evaluated [25]. Similarly, GDNF secreted by transplanted cells is considered to be one of the reasons that therapeutic effects are achieved by cell transplantation [26, 27]. Our group demonstrated that the implantation of encapsulated GDNF-secreting cells into 6-OHDA-lesioned striatum resulted in protective effects on dopaminergic neurons [28–30]. Rats receiving an implantation of GDNF-secreting cells at 2 weeks after 6-OHDA lesions were induced showed more neurorestorative effects compared to those receiving the implantation 4 weeks after 6-OHDA lesions were induced [29]. After this study was completed, we performed a new experiment using earlier timing for the implantation; that is, the implantation was per-

formed simultaneously with and at 1, 2, and 4 weeks after the 6-OHDA lesions were induced, with GFR $\alpha 1$ expression in the striatum and substantia nigra being evaluated at 1 and 2 weeks after the 6-OHDA lesions were induced [30]. In the study, the earlier implantation of GDNF resulted in a more improved behavioral score and better preserved dopaminergic neurons with a higher expression level of GFR $\alpha 1$ [30], thus revealing that GDNF-responsive fibers and neurons are necessary for GDNF therapy.

Advantages of GDNF therapy using encapsulated cell transplantation. Encapsulated cell transplantation has been clinically used for the treatment of diseases arising from hormone-producing organs like the pancreas and parathyroid [31, 32]. For the CNS diseases, encapsulated ciliary neurotrophic factor (CNTF)-producing cells and chromaffin cells were transplanted into ALS patients and patients with severe chronic pain, respectively [33, 34]. As described in the previous section, encapsulated cell transplantation was used to administer GDNF to a PD model of rats in our laboratory. Encapsulated cell transplantation has many advantages [35] (Fig. 1). 1) Various neurotransmitters or neurotrophic factors can be produced continuously from encapsulated cells with tailored properties. Cells inside the capsule survive with a sufficient supply of nutrients and oxygen through the semipermeable membrane and are capable of secreting factors outwards. 2) Few immune reactions and no immunological rejection arise because the cells inside are protected by a stiff envelope. The molecular sieve prevents immunocompetent cells from invading the capsule and attacking the donor cells. 3) Tumorigenesis does not occur because the donor cells stay inside the capsule. 4) The capsule can be removed from the transplanted brain when problems arise after transplantation. 5) Various cells including immortalized cell lines can be transplanted safely as surviving donors with almost no ethical problems. Compared to the infusion using a pump system, the delivery of freshly made factor with no degradation and the absence of a need for pump implantation are the advantages of encapsulated cell transplantation. A single administration of GDNF was useful for the PD model of rats [36], and delayed short-term GDNF administration close to the substantia nigra

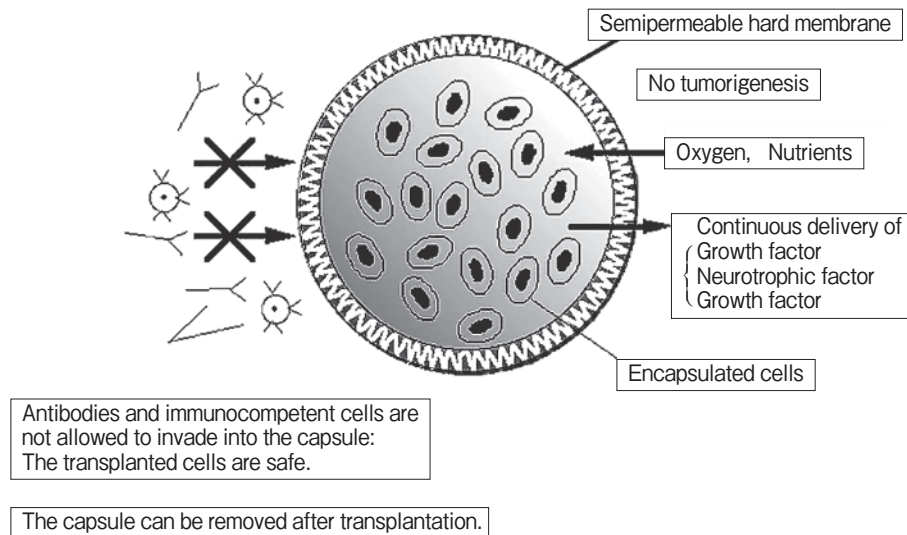


Fig. 1 Scheme of encapsulated cells. Capsules are made with semipermeable membrane. Oxygen and nutrition can freely move into the capsule, although antibodies and immunocompetent cells cannot pass through the membrane. In addition, the trophic factor or neurotransmitter secreted from cells inside the capsule can be expanded around the capsule.

protected against neuronal degeneration [37]. However, the study revealed that short-term GDNF administration could not achieve a long-term rescue of the nigral cells. Indeed, PD is slowly progressive, so a long-lasting effect is desirable. In our recent study, we used encapsulated cells secreting fresh GDNF at about 20 ng per day for 2 months *in vivo* [30]. Although the dose was relatively low compared to some studies, intraparenchymal continuous infusion appeared to be more effective than intraventricular infusion [38].

Current State of GDNF Therapy for PD Patients

Success of intraputaminial GDNF infusion in PD patients. Based on the successful results of the animal experiments described above, a clinical trial using GDNF was begun for PD patients. However, the initial several studies demonstrated that the intraventricular injection of GDNF did not improve the motor scores of PD patients and caused many side effects such as nausea, anorexia, vomiting, weight loss, hyponatremia, paresthesias including *Lehrmitte* signs and psychotic manifestations [39, 40]. The researchers considered that the unexpected poor results might be due to inappropriate GDNF

delivery, because intraputaminial GDNF infusion to aged non-human primates ameliorated the behavioral score and the metabolism of dopamine with no problematic side effects [41]. Dr. Gill and his colleagues then reported successful results using intraputaminial GDNF infusion in 2003 [42]. The direct administration of GDNF into the putamen of five PD patients improved the patients' Unified Parkinson's Disease Rating Scale (UPDRS) score in the off-medication state after 1 year with no serious side effects. Moreover, medication-induced dyskinesia was reduced by 64% and was not observed in the off-medication state during chronic GDNF delivery [42]. After this report, several groups demonstrated the therapeutic effect of GDNF on PD patients [43–45]. Dr. Gill's group in England continued GDNF infusion for 2 years and reported good outcomes with no side effects [44]. The same group demonstrated neuropathological evidence that GDNF infusion for 43 months caused sprouting of dopaminergic fibers in accordance with functional recovery [43]. In Kentucky, Dr. Slevin and his colleagues demonstrated that even unilateral GDNF infusion achieved bilateral improvement of motor functions with transient paresthesia in 20% of patients [45].

Recent controversy and hope surrounding GDNF therapy for PD patients. However in

2005, Amgen, the company holding the patents on GDNF therapy for PD, brought the clinical trial using GDNF to a halt, but not the pre-clinical and basic research [46, 47]. The main reasons for this decision were the negative results of the recent randomized controlled trial and the neutralizing anti-GDNF antibody found in 3 of 34 patients [48]. The latter was a factor even though these three patients were asymptomatic and the bioactivity and toxicity of the neutralizing antibody remains to be explored. In addition, cerebellar lesions were found in monkeys receiving a high dose of GDNF [46, 48]. These results and the decision of Amgen disappointed the patients and researchers involved in this study, and there has been fierce controversy over the decision [49–52]. However, it is too early to conclude that GDNF therapy is not feasible for PD patients, as such a conclusion would be based on just one study including several problems (an atypically younger patient population, and the GDNF dose and delivery system used in this study) as described in Dr. Baker's letter [53]. As many new experimental treatments are unlikely to be unqualified successes at first, we should re-consider the potential problems in the studies of GDNF therapy. If the administration of GDNF for a long time causes problems, we should investigate the long-term outcomes for patients receiving the GDNF infusion for just a few months. Alternatively, the effectiveness of intermittent administration of GDNF could be investigated. There are many questions to be answered before this research direction is abandoned. Recently, MRI analyses of nine PD patients with GDNF therapy revealed no cerebellar lesions in accord with a lack of cerebellar dysfunctions [54]. Continuous and steady efforts like the study from Kentucky are required, although the future of GDNF therapy is still unknown.

GDNF secreted from transplanted cells in PD patients. Recently the importance of transplanted cells as suppliers of trophic factors has come to light. In clinical application, autologous cells are relatively safe for patients and present almost no ethical issues; they are considered to be strong potential candidates for therapies for PD [55–57]. Dr. Arjona reported that 6 patients with advanced PD underwent bilateral transplantation of autologous carotid body cell aggregates into the striatum [55].

Bilateral intrastriatal transplantation was performed in six PD patients with both pre- and post-transplant (18 months) evaluations. No patients demonstrated side effects, including dyskinesia. In addition, 5 of 6 patients showed ameliorated UPDRSIII in their off-state. Carotid body-grafted Parkinsonian animals showed a nigrostriatal dopaminergic neurorestoration, at least partially resulting from GDNF secretion from the grafted cells [58]. In our laboratory, the therapeutic effects of genetically engineered GDNF-secreting neural stem cells are also being explored using a PD model of animals to enhance the therapeutic effects of the neural stem cell itself.

Conclusions

Key animal experiments, including our recent findings related to GDNF and clinical trials of GDNF therapy for PD patients, are described in this review. The future of GDNF therapy is uncertain, although GDNF has strong neuroprotective and neurorestorative effects. Steady and continuous efforts are required to elucidate the potencies of GDNF and its potential contribution to PD patients.

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References

1. Hornykiewicz O: Dopamine in the basal ganglia. Its role and therapeutic implications (including the clinical use of L-DOPA). *Br Med Bull* (1973) 29: 172–178.
2. Calne DB: Progress in Parkinson's disease. *N Engl J Med* (1984) 310: 523–524.
3. Lang AE and Lozano AM: Parkinson's disease. First of two parts. *N Engl J Med* (1998) 339: 1044–1053.
4. Lang AE and Lozano AM: Parkinson's disease. Second of two parts. *N Engl J Med* (1998) 339: 1130–1143.
5. Kumar R, Lozano AM, Kim YJ, Hutchison WD, Sime E, Hallett E and Lang AE: Double-blind evaluation of subthalamic nucleus deep brain stimulation in advanced Parkinson's disease. *Neurology* (1998) 51: 850–855.
6. Freed CR, Greene PE, Breeze RE, Tsai WY, DuMouchel W, Kao R, Dillon S, Winfield H, Culver S, Trojanowski JQ, Eidelberg D and Fahn S: Transplantation of embryonic dopamine neurons for severe Parkinson's disease. *N Engl J Med* (2001) 344: 710–719.
7. Olanow CW, Goetz CG, Kordower JH, Stoessl AJ, Sossi V, Brin MF, Shannon KM, Nauert GM, Peri DP, Godbold J and Freeman TB: A double-blind controlled trial of bilateral fetal nigral transplantation in Parkinson's disease. *Ann Neurol* (2003) 54: 403–414.

8. Takagi Y, Takahashi J, Saiki H, Morizane A, Hayashi T, Kishi Y, Fukuda H, Okamoto Y, Koyanagi M, Ideguchi M, Hayashi H, Imazato T, Kawasaki H, Suemori H, Omachi S, Iida H, Itoh N, Nakatsuji N, Sasai Y and Hashimoto N: Dopaminergic neurons generated from monkey embryonic stem cells function in a Parkinson primate model. *J Clin Invest* (2005) 115: 102–109.
9. Dezawa M, Kanno H, Hoshino M, Cho H, Matsumoto N, Itokazu Y, Tajima N, Yamada H, Sawada H, Ishikawa H, Mimura T, Kitada M, Suzuki Y and Ide C: Specific induction of neuronal cells from bone marrow stromal cells and application for autologous transplantation. *J Clin Invest* (2004) 113: 1701–1710.
10. Lin LF, Doherty DH, Lile JD, Bektesh S and Collins F: GDNF: a glial cell line-derived neurotrophic factor for midbrain dopaminergic neurons. *Science* (1993) 260: 1130–1132.
11. Kitagawa H, Hayashi T, Mitsumoto Y, Koga N, Itoyama Y and Abe K: Reduction of ischemic brain injury by topical application of glial cell line-derived neurotrophic factor after permanent middle cerebral artery occlusion in rats. *Stroke* (1998) 29: 1417–1422.
12. Li L, Wu W, Lin LF, Lei M, Oppenheim RW and Houenou LJ: Rescue of adult mouse motoneurons from injury-induced cell death by glial cell line-derived neurotrophic factor. *Proc of the Natl Acad of Sci USA* (1995) 92: 9771–9775.
13. Kim BT, Rao VL, Sailor KA, Bowen KK and Dempsey RJ: Protective effects of glial cell line-derived neurotrophic factor on hippocampal neurons after traumatic brain injury in rats. *J Neurosurg* (2001) 95: 674–679.
14. Tomac A, Lindqvist E, Lin LF, Ogren SO, Young D, Hoffer BJ and Olson L: Protection and repair of the nigrostriatal dopaminergic system by GDNF in vivo. *Nature* (1995) 373: 335–339.
15. Gash DM, Zhang Z, Ovadia A, Cass WA, Yi A, Simmerman L, Russell D, Martin D, Lapchak PA, Collins F, Hoffer BJ and Gerhardt GA: Functional recovery in parkinsonian monkeys treated with GDNF. *Nature* (1996) 380: 252–255.
16. Sagot Y, Tan SA, Hammang JP, Aebischer P and Kato AC: GDNF slows loss of motoneurons but not axonal degeneration or premature death of pmn/pmn mice. *J Neurosci* (1996) 16: 2335–2341.
17. Eggert K, Schlegel J, Oertel W, Wurz C, Krieg JC and Vedder H: Glial cell line-derived neurotrophic factor protects dopaminergic neurons from 6-hydroxydopamine toxicity in vitro. *Neurosci Lett* (1999) 269: 178–182.
18. Hebb AO, Hebb K, Ramachandran AC and Mendez I: Glial cell line-derived neurotrophic factor-supplemented hibernation of fetal ventral mesencephalic neurons for transplantation in Parkinson disease: long-term storage. *J Neurosurg* (2003) 98: 1078–1083.
19. Kirik D, Georgievska B, Rosenblad C and Bjorklund A: Delayed infusion of GDNF promotes recovery of motor function in the partial lesion model of Parkinson's disease. *Eur J Neurosci* (2001) 13: 1589–1599.
20. Kordower JH: In vivo gene delivery of glial cell line-derived neurotrophic factor for Parkinson's disease. *Ann Neurol* (2003) 53(Suppl 3): S120–S132; discussion S132–S134.
21. Treanor JJ, Goodman L, de Sauvage F, Stone DM, Poulsen KT, Beck CD, Gray C, Armanini MP, Pollock RA, Hefti F, Phillips HS, Goddard A, Moore MW, Buj-Bello A, Davies AM, Asai N, Takahashi M, Vandlen R, Henderson CE and Rosenthal A: Characterization of a multicomponent receptor for GDNF. *Nature* (1996) 382: 80–83.
22. Worby CA, Vega QC, Zhao Y, Chao HH, Seasholtz AF and Dixon JE: Glial cell line-derived neurotrophic factor signals through the RET receptor and activates mitogen-activated protein kinase. *J Biol Chem* (1996) 271: 23619–23622.
23. Nicole O, Ali C, Docagne F, Plawinski L, MacKenzie ET, Vivien D and Buisson A: Neuroprotection mediated by glial cell line-derived neurotrophic factor: involvement of a reduction of NMDA-induced calcium influx by the mitogen-activated protein kinase pathway. *J Neurosci* (2001) 21: 3024–3033.
24. Aoi M, Date I, Tomita S and Ohmoto T: Single or continuous injection of glial cell line-derived neurotrophic factor in the striatum induces recovery of the nigrostriatal dopaminergic system. *Neurol Res* (2000) 22: 832–836.
25. Yasuhara T, Matsukawa N, Hara K, Yu G, Xu L, Maki M, Kim SU and Borlongan CV: Transplantation of human neural stem cell exerts neuroprotection in a rat model of Parkinson's disease. *J Neurosci* (2006) 26: 12497–12511.
26. Chiang YH, Borlongan CV, Zhou FC, Hoffer BJ and Wang Y: Transplantation of fetal kidney cells: neuroprotection and neuroregeneration. *Cell Transplant* (2005) 14: 1–9.
27. Borlongan CV, Hadman M, Sanberg CD and Sanberg PR: Central nervous system entry of peripherally injected umbilical cord blood cells is not required for neuroprotection in stroke. *Stroke* (2004) 35: 2385–2389.
28. Date I, Shingo T, Yoshida H, Fujiwara K, Kobayashi K, Takeuchi A and Ohmoto T: Grafting of encapsulated genetically modified cells secreting GDNF into the striatum of parkinsonian model rats. *Cell Transplant* (2001) 10: 397–401.
29. Shingo T, Date I, Yoshida H and Ohmoto T: Neuroprotective and restorative effects of intrastriatal grafting of encapsulated GDNF-producing cells in a rat model of Parkinson's disease. *J Neurosci Res* (2002) 69: 946–954.
30. Yasuhara T, Shingo T, Muraoka K, Kobayashi K, Takeuchi A, Yano A, Wenji Y, Kameda M, Matsui T, Miyoshi Y and Date I: Early transplantation of an encapsulated glial cell line-derived neurotrophic factor-producing cell demonstrating strong neuroprotective effects in a rat model of Parkinson disease. *J Neurosurg* (2005) 102: 80–89.
31. Tibell A, Rafael E, Wennberg L, Nordenstrom J, Bergstrom M, Geller RL, Loudovaris T, Johnson RC, Brauker JH, Neuenfeldt S and Wernerson A: Survival of macroencapsulated allogeneic parathyroid tissue one year after transplantation in nonimmunosuppressed humans. *Cell Transplant* (2001) 10: 591–599.
32. Groth CG, Korsgren O, Tibell A, Tollemar J, Moller E, Bolinder J, Ostman J, Reinholt FP, Hellerstrom C and Andersson A: Transplantation of porcine fetal pancreas to diabetic patients. *Lancet* (1994) 344: 1402–1404.
33. Aebischer P, Schlupe M, Deglon N, Joseph JM, Hirt L, Heyd B, Goddard M, Hammang JP, Zurn AD, Kato AC, Regli F and Baetge EE: Intrathecal delivery of CNTF using encapsulated genetically modified xenogeneic cells in amyotrophic lateral sclerosis patients. *Nat Med* (1996) 2: 696–699.
34. Buchser E, Goddard M, Heyd B, Joseph JM, Favre J, de Tribolet N, Lysaght M and Aebischer P: Immunoisolated xenogenic chormaffin cell therapy for chronic pain. Initial clinical experience. *Anesthesiology* (1996) 85: 1005–1012; discussion 29A–30A.
35. Yasuhara T, Borlongan CV and Date I: Ex vivo gene therapy: transplantation of neurotrophic factor-secreting cells for cerebral ischemia. *Front Biosci* (2006) 11: 760–775.
36. Aoi M, Date I, Tomita S and Ohmoto T: Single administration of GDNF into the striatum induced protection and repair of the nigrostriatal dopaminergic system in the intrastriatal 6-hydroxydopamine injection model of hemiparkinsonism. *Restor Neurol Neurosci* (2001) 17: 31–38.

37. Winkler C, Sauer H, Lee CS and Bjorklund A: Short-term GDNF treatment provides long-term rescue of lesioned nigral dopaminergic neurons in a rat model of Parkinson's disease. *J Neurosci* (1996) 16: 7206-7215.
38. Aoi M, Date I, Tomita S and Ohmoto T: GDNF induces recovery of the nigrostriatal dopaminergic system in the rat brain following intracerebroventricular or intraparenchymal administration. *Acta Neurochir* (2000) 142: 805-810.
39. Nutt JG, Burchiel KJ, Comella CL, Jankovic J, Lang AE, Laws ER Jr., Lozano AM, Penn RD, Simpson RK Jr., Stacy M and Wooten GF: Randomized, double-blind trial of glial cell line-derived neurotrophic factor (GDNF) in PD. *Neurology* (2003) 60: 69-73.
40. Kordower JH, Palfi S, Chen EY, Ma SY, Sendera T, Cochran EJ, Mufson EJ, Penn R, Goetz CG and Comella CD: Clinicopathological findings following intraventricular glial-derived neurotrophic factor treatment in a patient with Parkinson's disease. *Ann Neurol* (1999) 46: 419-424.
41. Maswood N, Grondin R, Zhang Z, Stanford JA, Surgener SP, Gash DM and Gerhardt GA: Effects of chronic intraputamenal infusion of glial cell line-derived neurotrophic factor (GDNF) in aged Rhesus monkeys. *Neurobiol Aging* (2002) 23: 881-889.
42. Gill SS, Patel NK, Hottton GR, O'Sullivan K, McCarter R, Bunnage M, Brooks DJ, Svendsen CN and Heywood P: Direct brain infusion of glial cell line-derived neurotrophic factor in Parkinson disease. *Nat Med* (2003) 9: 589-595.
43. Love S, Plaha P, Patel NK, Hottton GR, Brooks DJ and Gill SS: Glial cell line-derived neurotrophic factor induces neuronal sprouting in human brain. *Nat Med* (2005) 11: 703-704.
44. Patel NK, Bunnage M, Plaha P, Svendsen CN, Heywood P and Gill SS: Intraputamenal infusion of glial cell line-derived neurotrophic factor in PD: a two-year outcome study. *Ann Neurol* (2005) 57: 298-302.
45. Slevin JT, Gerhardt GA, Smith CD, Gash DM, Kryscio R and Young B: Improvement of bilateral motor functions in patients with Parkinson disease through the unilateral intraputamenal infusion of glial cell line-derived neurotrophic factor. *J Neurosurg* (2005) 102: 216-222.
46. Peck P: Amgen decision to halt GDNF clinical trials and withdraw the drug triggers protest from researchers and patients. *Neurol Today: Am Acad Neurol* (2005) 5: 4, 7, 24.
47. Pollack A: Patients in test won't get drug, Amgen decides. *NY Times* (2005) 12: C1-2.
48. Lang AE, Gill S, Patel NK, Lozano A, Nutt JG, Penn R, Brooks DJ, Hottton G, Moro E, Heywood P, Brodsky MA, Burchiel K, Kelly P, Dalvi A, Scott B, Stacy M, Turner D, Wooten VG, Elias WJ, Laws ER, Dhawan V, Stoessl AJ, Matcham J, Coffey RJ and Traub M: Randomized controlled trial of intraputamenal glial cell line-derived neurotrophic factor infusion in Parkinson disease. *Ann Neurol* (2006) 59: 459-466.
49. The hard way to a Bill of Rights. *Lancet neurology* (2005) 4: 787.
50. Lang AE, Langston JW, Stoessl AJ, Brodsky M, Brooks DJ, Dhawan V, Elias WJ, Lozano AM, Moro E, Nutt JG, Stacy M, Turner D and Wooten GF: GDNF in treatment of Parkinson's disease: response to editorial. *Lancet neurology* (2006) 5: 200-202.
51. Penn RD, Dalvi A, Slevin J, Young B, Gash D, Gerhardt G and Hutchinson M: GDNF in treatment of Parkinson's disease: response to editorial. *Lancet neurology* (2006) 5: 202-203.
52. Slevin J, Gerhardt G, Smith CD, Gash D and Young AB: Reply: GDNF poses troubling questions for doctors, drug maker. *Ann Neurol* (2006) 59: A5-6.
53. Barker RA: Continuing trials of GDNF in Parkinson's disease. *Lancet neurology* (2006) 5: 285-286.
54. Chebrulu H, Slevin JT, Gash DA, Gerhardt GA, Young B, Given CA and Smith CD: MRI volumetric and intensity analysis of the cerebellum in Parkinson's disease patients infused with glial-derived neurotrophic factor (GDNF). *Exp Neurol* (2006) 198: 450-456.
55. Arjona V, Minguéz-Castellanos A, Montoro RJ, Ortega A, Escamilla F, Toledo-Aral JJ, Pardal R, Mendez-Ferrer S, Martin JM, Perez M, Katati MJ, Valencia E, Garcia T and Lopez-Barneo J: Autotransplantation of human carotid body cell aggregates for treatment of Parkinson's disease. *Neurosurgery* (2003) 53: 321-328; discussion 8-30.
56. Date I, Asari S and Ohmoto T: Two-year follow-up study of a patient with Parkinson's disease and severe motor fluctuations treated by co-grafts of adrenal medulla and peripheral nerve into bilateral caudate nuclei: case report. *Neurosurgery* (1995) 37: 515-518; discussion 8-9.
57. Itakura T, Komai N, Ryujin Y, Ooiwa Y, Nakai M and Yasui M: Autologous transplantation of the cervical sympathetic ganglion into the parkinsonian brain: case report. *Neurosurgery* (1994) 35: 155-157; discussion 7-8.
58. Toledo-Aral JJ, Mendez-Ferrer S, Pardal R, Echevarria M and Lopez-Barneo J: Trophic restoration of the nigrostriatal dopaminergic pathway in long-term carotid body-grafted parkinsonian rats. *J Neurosci* (2003) 23: 141-148.